

# Mercodia Ultrasensitive C-peptide ELISA

Directions for Use

**10-1141-01**

**REAGENTS FOR 96 DETERMINATIONS**

For *in vitro* diagnostic use



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Manufactured by

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## EXPLANATION OF SYMBOLS USED ON LABELS

	Reagents for 96 determinations
	Expiry date
	Store between 2–8°C
	Lot No.
	For <i>in vitro</i> diagnostic use

## INTENDED USE

Mercodia Ultrasensitive C-peptide ELISA provides a method for the quantitative determination of human C-peptide in serum, plasma or urine.

## SUMMARY AND EXPLANATION OF THE TEST

Qualitative and quantitative evaluation of pancreatic  $\beta$ -cell function is not only of use in the pre- and postdiagnostic study of the natural history of diabetes mellitus, but is also relevant in clinical practice as a guide to the correct choice of treatment. Peripheral insulin levels cannot be used to assess  $\beta$ -cell function because of a large and variable uptake from the portal circulation into the liver, and because insulin assays cannot distinguish endogenous from exogenous insulin.

Within the pancreatic  $\beta$ -cell, proinsulin is cleaved into one molecule of C-peptide and one molecule of insulin. C-peptide is subsequently released into the circulation at concentrations equimolar to those of insulin. In contrast to insulin, C-peptide is only minimally extracted by the liver. Peripheral C-peptide concentrations therefore reflect the secretion of  $\beta$ -cells more accurately than insulin.

## PRINCIPLE OF THE PROCEDURE

Mercodia Ultrasensitive C-peptide ELISA is a solid phase two-site enzyme immunoassay. It is based on the direct sandwich technique in which two monoclonal antibodies are directed against separate antigenic determinants on the C-peptide molecule. During incubation C-peptide in the sample reacts with anti-C-peptide antibodies bound to the microtitration well. After washing, peroxidase conjugated anti-C-peptide antibodies are added. After a second incubation and a simple washing step, the bound conjugate is detected by reaction with 3,3',5,5'-tetramethylbenzidine (TMB). The reaction is stopped by adding acid to give a colorimetric endpoint that is read spectrophotometrically.

## WARNINGS AND PRECAUTIONS

- For *in vitro* diagnostic use.
- The contents of this kit and their residues must not be allowed to come into contact with ruminating animals or swine.
- The Stop Solution in this kit contains 0.5 M  $\text{H}_2\text{SO}_4$ . Follow routine precautions for handling hazardous chemicals.
- All specimens should be handled as if capable of transmitting infections.

## MATERIAL REQUIRED BUT NOT PROVIDED

- Pipettes with appropriate volumes (repeating pipettes preferred for addition of enzyme conjugate solution, Assay Buffer, Substrate TMB and Stop Solution)
- Tubes, beakers and cylinders for reagent preparation
- Redistilled water
- Magnetic stirrer
- Vortex mixer
- Microplate reader with 450 nm filter
- Microplate shaker (700–900 cycles per minute, orbital movement)
- Microplate washing device with overflow function (recommended but not required)

## REAGENTS

Each Mercodia Ultrasensitive C-peptide ELISA (10-1141-01) kit contains reagents for 96 wells, sufficient for 42 samples and one calibrator curve in duplicate. For larger series of assays, use pooled reagents from packages bearing identical lot numbers. The expiry date for the complete kit is stated on the outer label. The recommended storage temperature is 2–8°C.

<b>Coated Plate</b> Mouse monoclonal anti-C-peptide	1 plate	96 wells 8-well strips	Ready for use
For unused microplate strips, reseal the bag using adhesive tape, store at 2–8°C and use within 8 weeks			
<b>Calibrators 1, 2, 3, 4, 5</b> Human C-peptide Color coded yellow Concentration stated on vial label Storage after reconstitution: 2–8°C for 1 week For storage of reconstituted Calibrators for more than 1 week, store at -20°C	5 vials	1000 µL	Lyophilized Add 1000 µL redistilled water per vial
<b>Calibrator 0</b> Color coded yellow	1 vial	5 mL	Ready for use
<b>Assay Buffer</b> Color coded red	1 vial	6 mL	Ready for use
<b>Enzyme Conjugate 11X</b> Peroxidase conjugated mouse monoclonal anti-C-peptide	1 vial	1.2 mL	Preparation, see below
<b>Enzyme Conjugate Buffer</b> Color coded blue	1 vial	12 mL	Ready for use
<b>Wash Buffer 21X</b> Storage after dilution: 2–8°C for 8 weeks	1 bottle	50 mL	Dilute with 1000 mL redistilled water to make wash buffer 1X solution
<b>Substrate TMB</b> Colorless solution <i>Note! Light sensitive!</i>	1 bottle	22 mL	Ready for use
<b>Stop Solution</b> 0.5 M H <sub>2</sub> SO <sub>4</sub>	1 vial	7 mL	Ready for use

## Preparation of enzyme conjugate 1X solution

Prepare the needed volume of enzyme conjugate 1X solution by dilution of Enzyme Conjugate 11X, (1+10) in Enzyme Conjugate Buffer or according to the table below. When preparing enzyme conjugate 1X solution for the whole plate, pour all of the Enzyme Conjugate Buffer into the Enzyme Conjugate 11X vial. Mix gently. Use within one day.

Number of strips	Enzyme Conjugate 11X	Enzyme Conjugate Buffer
12 strips	1 vial	1 vial
8 strips	700 µL	7 mL
4 strips	350 µL	3.5 mL

## SPECIMEN COLLECTION AND HANDLING

### Serum

Collect blood by venipuncture, allow to clot, and separate the serum by centrifugation. Samples can be stored at 2–8°C up to 3 days. For longer periods store samples at –20°C. Avoid repeated freezing and thawing.

### Plasma

Collect blood by venipuncture into tubes containing heparin or EDTA as anticoagulant, and separate the plasma fraction. Samples can be stored at 2–8°C up to 3 days. For longer periods store samples at –20°C. Avoid repeated freezing and thawing.

### Urine

Collect a 24 hour urine (without preservative). Keep the specimen at 2–8°C between collections. Record the total volume of the specimen and retain a well mixed aliquot for analysis. Store the samples at 2–8°C for a maximum of 24 hours before assay. For longer storage, keep the urine samples frozen at –70°C until assay is performed. Repeated freezing and thawing must be avoided. Cellular debris should be removed before assay, either by filtration or centrifugation.

## Preparation of samples

Urine. Dilute urine samples 1/10 v/v in Calibrator 0 before assay.

## TEST PROCEDURE

All reagents and samples must be brought to room temperature before use.

Prepare a calibrator curve for each assay run.

1. Prepare enzyme conjugate 1X solution and wash buffer 1X solution.
2. Prepare sufficient microplate wells to accommodate Calibrators, controls and samples in duplicate.
3. Pipette 25  $\mu$ L each of Calibrators, controls and samples into appropriate wells.
4. Add 50  $\mu$ L Assay Buffer to each well.
5. Incubate on a plate shaker (700-900 rpm) for 1 hour at room temperature (18–25°C).
6. Wash 6 times with 700  $\mu$ L wash buffer 1X solution per well using an automatic plate washer with overflow-wash function, after final wash, invert and tap the plate firmly against absorbent paper. Do not include soak step in washing procedure.  
Or manually,  
discard the reaction volume by inverting the microplate over a sink. Add 350  $\mu$ L wash buffer 1X solution to each well. Discard the wash solution, tap firmly several times against absorbent paper to remove excess liquid. Repeat 5 times. Avoid prolonged soaking during washing procedure.
7. Add 100  $\mu$ L enzyme conjugate 1X solution to each well.
8. Incubate on a plate shaker (700-900 rpm) for 1 hour at room temperature (18–25°C).
9. Wash as described in step 6.
10. Add 200  $\mu$ L Substrate TMB.
11. Incubate for 30 minutes on the bench at room temperature (18–25°C).
12. Add 50  $\mu$ L Stop Solution to each well.  
Place plate on a shaker for approximately 5 seconds to ensure mixing.
13. Read optical density at 450 nm and calculate results.  
Read within 30 minutes.

*Note!* To prevent contamination between the conjugate and substrate, separate pipettes are recommended.

## INTERNAL QUALITY CONTROL

Commercial controls and/or internal serum pools with low, intermediate and high C-peptide concentrations should routinely be assayed as samples, and results charted from day to day. It is good laboratory practice to record the following data for each assay: kit lot number, preparation dates of kit components, OD values for the blank, Calibrators and controls.

Laboratories should follow government regulations or accreditation requirements for quality control frequency.

## CALCULATION OF RESULTS

### Computerized calculation

The concentration of C-peptide is obtained by computerized data reduction of the absorbance for the Calibrators, except for Calibrator 0, versus the concentration using cubic spline regression.

### Manual calculation

1. Plot the absorbance values obtained for the Calibrators, except Calibrator 0, against the C-peptide concentration on a log-log paper and construct a calibrator curve.
2. Read the concentration of the samples from the calibrator curve.

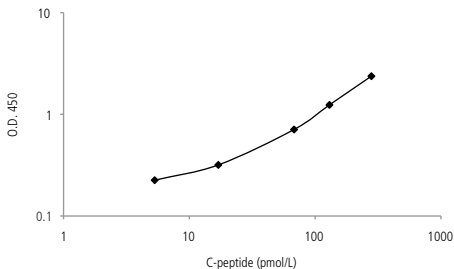
### Example of worksheet

Wells	Identity	A <sub>450</sub>	Mean conc. pmol/L
1A–B	Calibrator 0	0.086/0.090	
1C–D	Calibrator 1*	0.136/0.133	
1E–F	Calibrator 2*	0.237/0.237	
1G–H	Calibrator 3*	0.702/0.728	
2A–B	Calibrator 4*	1.405/1.419	
2C–D	Calibrator 5*	2.862/2.945	
2E–F	Sample 1	0.193/0.204	14
2G–H	Sample 2	0.374/0.391	35
3A–B	Sample 3	0.980/0.951	97

\*Concentration stated on vial label.

## Calibrator curve

A typical calibrator curve is shown here. Do not use this curve to determine actual assay results.



## LIMITATIONS OF THE PROCEDURE

As with all diagnostic tests, a definitive clinical diagnosis should not be based on the results of a single test, but should be made by the physician after all clinical findings have been evaluated.

Grossly lipemic, icteric or hemolyzed samples do not interfere in the assay.

## EXPECTED VALUES

Good practice dictates that each laboratory establishes its own expected range of values. The following results may serve as a guide until the laboratory has gathered sufficient data of its own.

Fasting serum levels for 136 tested, apparently healthy individuals, yielded a mean of 742 pmol/L (2.2 µg/L), a median of 628 pmol/L (1.9 µg/L) and a range, corresponding to the central 95% of the observations, of 343–1803 pmol/L (1.0–5.4 µg/L).



## PERFORMANCE CHARACTERISTICS

### Detection limit

Detection limit is defined as the Capability of Detection according to ISO11843-Part 1. Capability of Detection should be seen as part of a method validation, rather than the lowest concentration that can be measured.

The detection limit is  $\leq 2.5$  pmol/L (0.0076  $\mu\text{g/L}$ ) as determined by the methodology described in ISO11843- Part 4.

Concentration of samples with absorbance below Calibrator 1 should not be calculated, instead expressed as less or equal to ( $\leq$ ) the concentration indicated on the vial for Calibrator 1.

### Recovery

Serum:

Recovery upon addition is 93–103% (mean 99%).

Recovery upon dilution is 92–111% (mean 101%)

### Hook effect

Samples with a concentration of up to 36 nmol/L can be measured without giving falsely low results.

### Precision

Each sample was analyzed in 4 replicates on 5 different occasions.

Sample	Mean value pmol/L	Coefficient of variation		
		within assay %	between assay %	total assay %
1	37	3.8	5.5	6.7
2	60	2.6	4.2	5.0
3	143	2.5	1.6	2.9

### Specificity

Proinsulin at a concentration of 826 pmol/L yielded a result below the detection limit. For other peptides, the following cross reactions have been found:

Insulin	<0.0006%
Proinsulin	<1.8%
Proinsulin Des (31–32)	3 %
Proinsulin Split (32–33)	2 %
Proinsulin Des (64–65)	74 %
Proinsulin Split (65–66)	10 %

## **CALIBRATION**

Mercodia Ultrasensitive C-peptide ELISA kit is calibrated against the International Reference Reagent for C-peptide, IRR C-peptide 84/510.

## **CONVERSION FACTOR**

1 µg/L corresponds to 331 pmol/L

## **WARRANTY**

The performance data presented here was obtained using the procedure indicated. Any change or modification in the procedure not recommended by Mercodia AB may affect the results, in which event Mercodia AB disclaims all warranties expressed, implied or statutory, including the implied warranty of merchantability and fitness for use.

Mercodia AB and its authorised distributors, in such event, shall not be liable for damages indirect or consequential.

## **REFERENCES**

Craig ME, Howard NJ, Silink M, Rawlinson WD (2003) Reduced frequency of HLA DRB1\*03-DQB1\*02 in children with type 1 diabetes associated with enterovirus RNA. *J Infect Dis* 187:1562-1570

Gaines-Das RE and Bristow AF (1988) WHO International reference reagents for human proinsulin and human C-peptide. *J Biol Stand* 16:179-186

Further references can be found on our website: [www.mercodia.com](http://www.mercodia.com)



**SUMMARY PROTOCOL SHEET****Mercodia Ultrasensitive C-peptide ELISA**

Add Calibrators, controls* and samples	25 µL
Add Assay Buffer	50 µL
Incubate	1 hour at 18–25°C on a shaker 700-900 rpm
Wash with wash buffer 1X solution	700 µL, 6 times
Add enzyme conjugate 1X solution	100 µL
Incubate	1 hour at 18–25°C on a shaker 700-900 rpm
Wash with wash buffer 1X solution	700 µL, 6 times
Add Substrate TMB	200 µL
Incubate	30 minutes at 18-25°C
Add Stop Solution	50 µL Shake for 5 sec to ensure mixing
Measure A <sub>450</sub>	Evaluate results

\*not provided

For full details see page 6